



Comparison of OxOral® and NaOCl irrigants efficiency in *Enterococcus faecalis* elimination

Comparación de la eficacia de los irrigantes OxOral® y NaOCl en la eliminación de *Enterococcus faecalis*

Arely Herrera Saucedo,* Marco Antonio Corona Guerra,[§] Francisco Javier Vara Padilla,[§]
Dulce Haydeé Gutiérrez Valdez,^{||} Sandra Laura Alavez Rebollo[¶]

ABSTRACT

Objective: To compare effectiveness of OxOral® versus sodium hypochlorite in *Enterococcus faecalis* elimination at 15 and 60 seconds. **Material and methods:** Material used in the study was 36 *E. faecalis* ATCC 29212 cultures assigned to two groups: OxOral® and 5.25% sodium hypochlorite. Both groups were in turn divided into 15 and 60 second samples. Samples were placed in peptone water, 1 mL of irrigating solution and 1 mL of strain were left to rest. 1 mL was extracted at each time, samples were seeded into blood agar for 24 hours. Mann-Whitney U test was applied. **Results:** With sodium hypochlorite at 15 seconds, there were three cultures with acceptable growth and six with extended growth; at 60 seconds four cultures exhibited effective result, three acceptable and one extended. With OxOral® there was extended growth in all nine cultures at both established times, significant statistical differences were found at the 60 seconds time ($p < 0.01$). **Conclusion:** *E. faecalis* elimination was better with sodium hypochlorite at 60 seconds.

Key words: *E. faecalis*, sodium hypochlorite, OxOral®, effectiveness.
Palabras clave: *E. faecalis*, hipoclorito de sodio, OxOral®, eficacia.

INTRODUCTION

Root canal treatment targets elimination of injured pulp tissue, bacteriae and their endotoxins. To achieve this the following is needed: irrigation, biomechanical preparation (facilitating disposal of organic tissue) and sealing of root canals so as to prevent their subsequent contamination.¹⁻⁴ Although biomechanical preparation significantly reduces microbiota, it doesn't fully eliminate bacteriae in lateral canals, accessory canals, isthmus and apical delta; therefore selection of irrigating solution and medication to use within the canal will be of the utmost importance in order to reach areas not accessible during instrumentation; suitable apical sealing is equally important.⁵⁻⁸

One of the main causes of canal treatment failure is persistent multiplication and migration of bacteriae within the canals toward tissues around the root, caused

RESUMEN

Objetivo: Comparar la eficacia en la eliminación de *Enterococcus faecalis* con OxOral® versus hipoclorito de sodio a los 15 y 60 segundos. **Material y métodos:** Se incluyeron 36 cultivos de *E. faecalis* ATCC 29212 asignados en dos grupos; OxOral® e hipoclorito de sodio al 5.25% que a su vez fueron divididos en 15 y 60 segundos. Se colocaron 8 mL de agua peptonada, 1 mL del irrigante y 1 mL de la cepa, se dejó reposar. A cada tiempo se extrajo 1 mL y se sembró en agar sangre por 24 horas. Se empleó U de Mann-Whitney. **Resultados:** Con hipoclorito de sodio a 15 segundos hubo tres cultivos con crecimiento aceptable y seis extendido; a los 60 segundos, cuatro tuvieron resultado eficaz, tres aceptable, uno extendido. Con OxOral® hubo crecimiento extendido en los nueve cultivos, en ambos tiempos, encontrando diferencias estadísticamente significativas a los 60 segundos ($p < 0.01$). **Conclusión:** La eliminación de *E. faecalis* fue mejor con hipoclorito de sodio a los 60 segundos.

by deficient chemical and mechanical preparation. Adherence to dentin is the first step for bacterial colonization; later, invasion towards dentin tubules and biofilm formation take place.^{9,10} *Enterococcus*

* Endodontics Graduate Student.

§ Endodontics Professor, School of Dentistry.

|| Teacher/Researcher, School of Dentistry.

¶ Basic Area Coordinator, School of Dentistry.

Mexico Technological University, UNITEC.

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faecalis is within the range of microorganisms that can be found, this strain is found in 33% of all cases requiring a second root canal treatment,⁸ with peri-radicular lesions that were not repaired.⁵ This bacteria has additionally been associated to caries lesions, chronic periodontitis and persisting apical periodontitis.^{11,12} It possesses the ability to adapt to different circumstances (nutrient shortages, acidity, heat, alkalinity, ultraviolet light), thus it can effectively remain in medicated canals.^{11,13}

Its gelatinase activity contributes to its long term survival within filled canals^{13,14} favoring bonding with dentin; irrigating solutions contribute to eliminate bacteriae located in dentin canals or tubules.¹⁵

There are many types of irrigating solutions available in the market. Such is the case of sodium hypochlorite (NaOCl) hydrogen peroxide, chlorhexidine, EDTA (ethylenediaminetetraacetic acid) and superoxidation electrolyzed solution (OxOral®). Sodium hypochlorite, at concentrations of 0.5 to 5.25% possesses the capacity of eliminating organic residues in areas that instruments cannot reach, it therefore constitutes a suitable anti-microbial agent^{3,8,16} with tissue dissolution ability.^{6,7} One of its disadvantages is its toxicity on tissues surrounding the root, it can elicit pain, bleeding, volume increase, inflammation and tissue necrosis.⁷

Superoxidation electrolyzed solution (OxOral®) exerts disinfectant and sterilizing activity; this is due to its activity on bacteriae, viruses, fungi, spores and its low toxicity on tissues.¹⁷ These are electrochemically processed solutions, made from pure water and salt, which induce formation of elements derived from oxygen, hydrogen and chlorine, they are purified through inverse osmosis incorporating sodium chloride under voltage and current parameters to obtain ions and free radicals.¹⁷⁻²⁰

Among its antimicrobial properties we can mention, among others, activity against *Enterococcus faecalis*.^{17,18} Due to its recent appearance in the market, there are yet few literature reports on its bactericidal effect, existing reports show nil effect on *E. faecalis*.²¹

Comparison between super oxidant electrolyzed solution (OxOral®) and NaOCl has not been widely established; no precise benefits have been demonstrated in order to stop replication of microorganisms and in lesser time of use.

The target of the present study was to compare effectiveness of two irrigating solutions: super oxidant electrolyze solution (OxOral®) and NaOCl in the elimination of *E. faecalis* at two different times for each irrigating solution (15 and 60 seconds).

MATERIAL AND METHODS

An *in vitro* experimental study was conducted at the School of Dentistry of the Technological University of Mexico in the timeframe of August-December 2014. The study comprised 36 culture samples of *Enterococcus faecalis* TCC 29212, assigned to two groups: one for OxOral® and the other to 5.25% sodium hypochlorite; samples were studied at two different times: 15 and 60 seconds.

For recuperation and confirmation purposes, the strain was inoculated in a Petri dish with ram's blood agar at 5%, sowing in stretches for 24 hours at 37.5 °C. Once strain growth was obtained, a well isolated colony was harvested with a sterile handle, touching its upper section; it was then transferred to a test tube with 10 mL peptone water. 1 mL of aliquot was placed. In another test tube with 9 mL peptone water; this process was repeated until obtaining a second tube with lesser turbidity. One ml aliquot of this second tube was taken to be then mixed with 8 mL peptone water and 1 mL disinfectant. It was shaken and then left to rest for 15 and 60 seconds for both disinfectant solutions respectively.

At both times, 1 mL was extracted and seeded in Petri dish by extension in ram blood agar at 5%, and left to incubate for 24 hours at 37.5 °C.

After this, colony forming units (CFU) were counted. Boxes with zero to one colony indicated disinfectant maximum effectiveness. *E. faecalis* elimination was determined as acceptable (CFU still controlled by the disinfectant from 2 to 100) extended (uncontrolled CFU growth ; over 100) and effective (no CFU). Information was analyzed with program SPSS 17.0; irrigating solution effects were compared with Mann-Whitney U test.

RESULTS

It was observed that at 15 seconds, with OxOral® there was extended growth in all nine cultures; with sodium hypochlorite, acceptable growth was observed in three cultures and extended growth was seen in six cultures (*Figure 1*).

At 60 seconds extended growth was observed in all cultures with OxOral®; with sodium hypochlorite, effective result was found in four cultures, three of them with acceptable growth, one with extended growth and one culture could not be assessed due to processing error (*Figure 2*).

No statistically significant differences were found at 15 seconds ($p = 0.065$), it can thus be concluded that no significant bactericidal ability to eliminate *Enterococcus faecalis* was exhibited by NaOCl and

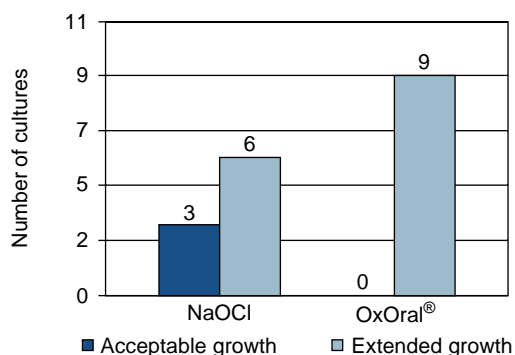


Figure 1. Comparison of *E. faecalis* growth in Petri dish after 15 seconds.

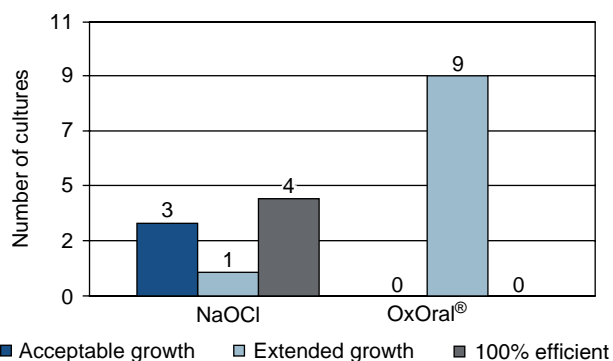


Figure 2. Comparison of *E. faecalis* growth in Petri dish after 60 seconds.

OxOral® when used for 15 seconds. Nevertheless, when used for 60 seconds statistical significant differences were found ($p < 0.01$) (the case with non evaluated sample was eliminated from the analysis). In the elimination process of *E. faecalis*, NaOCl at 5% proved to be the most effective when used for 60 seconds.

DISCUSSION

There are many studies reporting effectiveness of hypochlorite (2.5 to 6%), for this reason, it is used as a comparison point with other irrigating solutions that appear in the market, such is the case of OxOral® whose effectiveness has not been fully studied.

Cobankara⁷ reports on 5.25 NaOCl effectiveness and chlorine dioxide (ClO_2) in organic tissue dissolution, nevertheless, bacterial content was not analyzed; Wang¹⁶ reported that 6% NaOCl exhibited strongest antibacterial activity. In the present study, use of 5.25% NaOCl for 15 seconds

revealed extended growth of *E. faecalis* in six out of nine cultures, that is to say at this time, CFU growth was not controlled by the disinfectant. Nevertheless it was found to be more effective when the time was 60 seconds, since only one culture exhibited growth. Results in the present study concur with those of Gutmann⁶ who mentions that in order to obtain NaOCl effectiveness as bacterial control and tissue dissolution, it is necessary to work with 2.5 to 6% concentrations. This is similar to Harrison's reports²² of a study effected to verify 2.62 and 5.25% NaOCl antimicrobial effectiveness at period ranging from 15 to 120 seconds in cones contaminated with *E. faecalis*; he mentioned that after 45 seconds at a 5.25% concentration, and after 60 seconds at 2.62% concentration no *E. faecalis* bacterial growth could be observed. On the other hand, Souza²³ analyzed antimicrobial activity of NaOCl at different concentrations (1%, 0.5%, 0.12% and 0.25%) in paper cones previously contaminated with *E. faecalis*. Results indicated that it was eliminated at 0.5 and 1% concentrations during 15 seconds, this not being the case for the other concentrations.

When OxOral® was left in place for 15 and 60 seconds, CFU extended growth was found in all analyzed cultures, therefore, no CFU growth control could be reported, that is to say it did not exhibit bactericidal or bacteriostatic capacity against *E. faecalis*. Rojas²¹ conducted an *in vitro* study on contaminated files of teeth inoculated with the strain; in said study he concurred with our results and mentioned that OxOral® effected nil bactericidal effect on *E. faecalis* after 15 minutes, as well as after a 72 hour period; in cases when it has previously been used he mentioned that this solution was not effective for sterilization of endodontic instruments infected with *E. faecalis*.

Meanwhile, Zaragoza²⁴ conducted a study to compare antimicrobial effect of OxOral® Sterilizing and ACCUA Aséptico Hp®. To this effect he used strains of *S. aureus*, *S. mutans*, *L. acidophilus*, *C. albicans* and *E. coli* and *Pseudomona sp.* He mentioned that no inhibition was found when using OxOral®, therefore it can be concluded that the solution did not meet with properties pertaining to a sterilizing agent.

CONCLUSION

In the present study it was found that 5.25% NaOCl for 60 seconds was the best disinfectant to eliminate *E. faecalis*. This was not the case for OxOral® which exhibited growth in all cultures.

REFERENCES

1. Walton RE, Torabinejad M. *Endodoncia principios y práctica*. 2a edición. México: Editorial McGraw-Hill Interamericana; 1997. p. 1.
2. Cohen S. *Vías de la pulpa*. 8a edición. España: Editorial Elsevier Science; 2004. p. 108.
3. Thomas JE, Sem DS. An *in vitro* spectroscopic analysis to determine whether para-chloroaniline is produced from mixing sodium hypochlorite and chlorhexidine. *J Endod*. 2010; 36 (2): 315-317.
4. Delgado RJ, Gasparoto TH, Sipert CR, Pinheiro CR, Moraes IG, Garcia RB et al. Antimicrobial effects of calcium hydroxide and chlorhexidine on *Enterococcus faecalis*. *J Endod*. 2010; 36 (8): 1389-1393.
5. Arens DE. *Practical lessons in endodontic treatment*. China: Editorial Quintessence Publishing; 2009. pp. 145-146.
6. Gutmann JL. *Solución de problemas en endodoncia, prevención, identificación y tratamiento*. 4a edición. España: Editorial Elsevier Science; 2007. pp. 3-114.
7. Cobankara FK, Ozkan HB, Terlemez A. Comparison of organic tissue dissolution capacities of sodium hypochlorite and chlorine dioxide. *J Endod*. 2010; 36 (2): 272-274.
8. Ballal NV, Moorkoth S, Mala K, Bhat KS, Hussien SS, Pathak S. Evaluation of chemical interactions of maleic acid with sodium hypochlorite and chlorhexidine gluconate. *J Endod*. 2011; 37 (10): 1402-1405.
9. Kandaswamy D, Venkateshbabu N. Root canal irrigants. *J Conserv Dent*. 2010; 13 (4): 256-264.
10. Canalda C, Brau E. *Endodoncia técnicas clínicas y bases científicas*. España: Editorial Masson; 2001. pp. 29-36.
11. Al-Ahmad A, Müller N, Wiedmann-Al-Ahmad M, Sava I, Hübner J, Follo M et al. Endodontic and salivary isolates of *Enterococcus faecalis* integrate into biofilm from human salivary bacteria cultivated *in vitro*. *J Endod*. 2009; 35 (7): 986-991.
12. Zhu X, Wang Q, Zhang C, Cheung GS, Shen Y. Prevalence, phenotype, and genotype of *Enterococcus faecalis* isolated from saliva and root canals in patients with persistent apical periodontitis. *J Endod*. 2010; 36 (12): 1950-1955.
13. Madhubala MM, Srinivasan N, Ahamed S. Comparative evaluation of propolis and triantibiotic mixture as an intracanal medicament against *Enterococcus faecalis*. *J Endod*. 2011; 37 (9): 1287-1289.
14. Lovato KF, Sedgley CM. Antibacterial activity of endosequence root repair material and proroot MTA against clinical isolates of *Enterococcus faecalis*. *J Endod*. 2011; 37 (11): 1542-1546.
15. Cohen SK, Hargreaves KM. *Vías de la pulpa*. 9a edición. España: Editorial Elsevier Science; 2009. p. 347.
16. Wang Z, Shen Y, Haapasalo M. Effect of smear layer against disinfection protocols on *Enterococcus faecalis*-infected dentin. *J Endod*. 2013; 39 (11): 1395-1400.
17. Durán-Vega HC. Soluciones de superoxidación y su evolución tecnológica. *Dol Foro Nal Invest Clín Méd*. 2010; 7 (3): 4-8.
18. Pérez-Romano L, González-Espinosa D, Nuñez-Ochoa L, Landa-Solís C, Gutiérrez AA. *Solución de super-oxidación Microdacyn 60MR*. Una tecnología de vanguardia para tratar heridas. Instituto Nacional de Rehabilitación, Secretaría de Salud de México, Facultad de Medicina y Veterinaria, Universidad Nacional Autónoma de México; 2005.
19. Rodríguez-Gorozpe CI, Jácome Musule JL, Perea-Mejía LM. Estudio comparativo de filtración microbiana coronal con tres diferentes materiales de restauración provisional en dientes obturados con Guttaflow. *Rev Odont Mex*. 2010; 14 (1): 21-31.
20. Bergenholts G, Horsted-Bindslev P, Reit C. *Text of endodontology*. Reino Unido: Editorial Blackwell Munksgaard; 2003. p. 308.
21. Rojas-Briones ME, Silva-Herzog Flores D, González-Amaro AM, Oliva-Rodríguez R. Evaluación comparativa de la capacidad antimicrobiana de una solución electrolizada de superoxidación con pH neutro y una solución a base de peróxido de hidrógeno. *Rev ADM*. 2013; 70 (4): 183-189.
22. Harrison JW, Wagner GW, Henry CA. Comparison of the antimicrobial effectiveness of regular and fresh scent Clorox. *J Endod*. 1990; 16 (7): 328-330.
23. Monteiro-Souza M, Gugelmin MCM, Saquy PC, Pécora JD. Ação antimicrobiana do hipoclorito de sódio em diferentes concentrações e tempos de contato. *Odonto*. 1992; 2 (4): 302-306.
24. Zaragoza-Meneses MTJ, López-Badillo LE, Rodríguez-Martínez D. Comparación del efecto antimicrobiano de dos soluciones esterilizantes de superoxidación con pH neutro. *Odont Act*. 2012; 9 (110): 38-40.

Mailing address:
Sandra Laura Alavez Rebollo
 E-mail: sandra_biol_uni@hotmail.com